## Furan- and Triazolinedione-based Chemistries for Bio-orthogonal Protein Modification

## Prof. Annemieke Madder

Organic and Biomimetic Chemistry Research (OBCR) group, Department of Organic and Macromolecular Chemistry Ghent University, Krijgslaan 281, S4, 9000 Ghent, Belgium annemieke.madder@ugent.be

Within OBCR, we have developed a highly selective and efficient singlet oxygen mediated crosslink technology which is applicable to peptide-protein, peptide-nucleic acid and nucleic acid interstrand crosslink scenarios.<sup>[1]</sup> For this purpose, a furan 'warhead' is introduced into one of the biomolecular partners and subsequently activated by means of an oxidation trigger such as singlet oxygen which induces generation of a nucleophile-sensitive keto-enal moiety.<sup>[2]</sup> The overall procedure allows spatiotemporal control of the crosslinking event.

In the context of peptide ligand-receptor interactions, we have described, in live cells under normal growth conditions, spontaneous enzymatic activation and crosslinking of furan-modified peptide ligands to their membrane GPCR with zero toxicity, high efficiency and spatio-specificity.<sup>[3]</sup> Furan introduction into peptide and protein ligands and subsequent covalent modification of their natural targets was achieved after triggering photocatalytic singlet oxygen generation.<sup>[4]</sup>

Different furan<sup>[5]</sup> and triazolinedione<sup>[6]</sup> based building blocks were further developed for versatile and site-selective modification of proteins and synthesis of bioconjugates. The talk will highlight selected specific examples of these cross-linking and conjugation methodologies.

The work was supported by the FWO-Vlaanderen, the BOF-UGent, the IOF-UGent and the European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant agreement No 721613 (MMBio ITN), No 956070 (OligoMed ITN), No. 665501 (Pegasus<sup>2</sup> fellowship).

## **References:**

<sup>1</sup> a) Carrette, L.L.G, Morii, T.; Madder, A. *Bioconj. Chem.* **2013**, *24(12)*, 2008-2014; b) L. L. G. Carrette, E. Gyssels, N. De Laet and A. Madder. *Chem Comm.* **2016**, 52, 1539.

<sup>2</sup> a) Op de Beeck, M., and Madder, A. JACS **2012**, *134*, 10737–10740; b) De Laet, N., Llamas, E.M., and Madder, A. *ChemPhotoChem* **2018** *2*, 575–579.

<sup>3</sup> a) Vannecke, Van Troys, Ampe & Madder, ACS Chemical Biology **2017**, 2191; b) EP10196898.0.; c) EP 15176415.6. <sup>4</sup> a) Miret-Casals, L.; Vannecke, W.; Hoogewijs, Madder, A. *et al. Chem. Comm.*, **2021**, **57**, 6054 – 6057; b) Miret Casals, L.; Van De Putte, S.; Madder, A. *et al.* Frontiers in Chemistry, **2022.** 9:799706.

<sup>5</sup> De Geyter, E.; Antonatou, E.; Kalaitzakis, D.; Madder, A. *Chemical Science*, **2021**, **12**, 5246 – 5252. EP 19160048.5.

<sup>6</sup>a) Decoene, K. W.; Ünal, K.; Staes, A.; Zwaenepoel, O.; Gettemans, J.; Gevaert, K.; Winne, J. M.; Madder, A. *Chemical Science*, **2022**, **13**, 5390 – 5397.